1 Introduction

There exist several techniques for imaging spatiotemporal distributions of mechanical properties in biological tissues and engineered constructs on scales from molecules to organs. Collectively, they are known as elasticity imaging. Diagnostic techniques employ phase-sensitive imaging modalities capable of tracking local tissue movements induced by a mechanical stimulus. The resulting image displays components of displacement or strain and sometimes a compliance or modulus. For example, ultrasonic and magnetic resonance (MR) techniques are frequently applied to breast tissues to image viscoelastic properties of tumors [1–3]. The principal advantage of elasticity imaging is the large object contrast for tissue stiffness [4] that occurs within stromal tissues in response to the advancing disease [5,6]. Another large application area is vascular elasticity imaging using MR [7], optical [8], x-ray [9], and ultrasonic [10] methods. Emerging applications include viscoelastic imaging of macromolecules [11] and engineered tissue constructs [12]. The excitement about elasticity imaging is extending beyond diagnosis as we increase our understanding of the role of cellular mechanochemical transduction [13], particularly in cancer [5] and atherosclerosis [14].

Clinical elasticity imaging of breast cancer patients shows that malignant tumors most frequently appear as stiff regions (low strain or high modulus) compared to background media [15,16]. Stiffening is common because of edema, cellular hyperplasia, and characteristic increases in stromal collagen concentration and cross-linking. However, cancers can also appear softer than the background tissue [17] because the magnitude, spatial homogeneity, and temporal variation of the strain response depend on the physiology [18] and tumor microenvironment [6] of a specific patient. In addition, images of viscoelastic properties show both lower [19] and higher [2,3] response times for malignant masses as compared to benign masses. Although electron microscopy data show changes in the connective tissue ultrastructure [20] that suggest lower viscosity, not enough is known about the viscoelastic behavior of breast tissues in vivo to determine if the diversity of findings are due to patient or experimental variabilities. To advance diagnostic applications, we must discover how disease-related changes to molecular bonding within stromal tissues affect the broad spectrum of viscoelastic responses. This is essentially the inverse problem of estimating structural features of polymers from measured mechanical properties.

This paper reviews classical linear theory for polymers undergoing standard mechanical (quasi-static) stimuli in the context of ultrasonic strain imaging. We investigate the role of discrete rheological models (Voigt and Maxwell) that offer concise parametric summaries of viscoelastic behavior. Measurements of gelatin gels with different experimental geometries test the validity of model assumptions, show the consequences of violations, and define ultrasonic imaging parameters required for strain imaging. Gelatin shares a basic structure and many features of stromal breast tissues, and yet it is a simpler medium with adjustable mechanical properties. Therefore, gelatin gels are excellent media for investigating the strengths and weaknesses of elasticity imaging. One long-term goal of elasticity imaging research is to interpret microstructural reorganization of connective tissues during cancer progression from the macroscopic deformation patterns in viscoelastic images. Our experience with gelatin provides a framework for future tissue investigations.

2 Methods

This section reviews constitutive equations for the experimental geometries used in this study, including strain imaging where stress and strain vary in space and time. Imaging techniques often
apply stresses and measure time-varying strain patterns; therefore, the discussion is focused on creep. Results from other geometries and stimuli allow comparisons for validating imaging techniques.

2.1 Constitutive Equation. Assume a small cubic volume of gelatin is centered at vector position \( x \). Applying a weak force to volume surfaces at \( t = t_0 \) produces infinitesimal stresses \( d\sigma_j(x,t') \), where \( t' = (t-t_0) \). These induce infinitesimal strains \( d\epsilon_j(x,t) = S_{ijkl} \epsilon_{ij}(x,t') d\sigma_l(x,t') \) for \( t' > 0 \), where the material properties of the medium are elements of the fourth-order compliance tensor \( S_{ijkl} \). In media with linear time-invariant material properties, strains histories can be superimposed [21,22] to find

\[
\epsilon_j(x,t) = \int_0^t dt' S_{ijkl}(x,t-t') \frac{d\sigma_l(x,t')}{dt'}
\]

Equation (1) describes time-varying strain for volume elements within a linear viscoelastic medium, and thus it also describes the strain image of a deformed object.

Adopting the notation \( \epsilon(x,t) = L \epsilon(x,t) = \int_0^t dt \exp(-\epsilon t) \epsilon(x,t) \) for the one-sided Laplace transform, Eq. (1) becomes

\[
\tilde{\epsilon}_j(x,s) = s \tilde{S}_{ijkl}(x,s) \tilde{\sigma}_l(x,s)
\]

Here, \( s \) is a complex variable fundamental to the Laplace transform. For isotropic media, \( \tilde{S} \) can be expanded to give the generalized viscoelastic Hooke’s law (cf. Eq. (11.2-8) [23])

\[
\tilde{\epsilon}_j(x,s) = \left( \frac{1}{9} \tilde{B}(x,s) - \frac{1}{6} \tilde{J}(x,s) \right) \tilde{\sigma}_i(x) \delta_{ij} + \frac{1}{3} \tilde{J}(x,s) \tilde{\sigma}_i(x)
\]

where \( \tilde{B}(x,s) = \tilde{\sigma}_{11}(x,s) + \tilde{\sigma}_{22}(x,s) + \tilde{\sigma}_{33}(x,s) \) is the trace of the stress matrix and \( \delta_{ij} \) is the Kronecker delta. \( \tilde{B}(x,s) \) is bulk compliance that describes volume changes in the medium and \( \tilde{J}(x,s) \) is shear compliance that describes shape changes, both in the Laplace domain. The subscripts \( kl, ij \) are interchangeable since the stress and strain tensors are the same size and have only six independent terms.

The task now is to formulate stress tensors for different measurement conditions and apply Eq. (3) to predict strain. In this manner, the results of standard measurement techniques with known geometry can be compared to those of imaging experiments where the geometry is less well known.

2.2 Uniaxial Compressive Stress: Creep. Our imaging experiments involve application of a uniaxial compressive stress under free-slip boundary conditions. Ideally, this experiment generates only one nonzero stress element, i.e., \( \tilde{\sigma}_{11} \), and three normal strains, although \( \tilde{\epsilon}_{22} = \tilde{\epsilon}_{33} \) for isotropic materials. Solving Eq. (3) for the strain tensor corresponding to the applied stress yields

\[
\tilde{\epsilon}_{11}(x,s) = \left( \frac{1}{9} \tilde{B}(x,s) + \frac{1}{3} \tilde{J}(x,s) \right) \tilde{\sigma}_{11}(x,s)
\]

For ultrasonic strain imaging, strain is estimated along the axis of the sound beam and in the direction of the applied force \( s_1 \). Consequently, \( \tilde{\epsilon}_{11} \) in Eq. (4) is often referred to as axial strain in imaging experiments [24]. Axial strain images are common because ultrasonic echoes are most sensitive to object movements along the phase-sensitive beam axis. In the following, \( \tilde{e} \) indicates \( \tilde{\epsilon}_{11} \) except where otherwise noted.

From one strain measurement, however, only the linear combination of shear and bulk compliances can be determined. Thus, we study the measurable quantity compressive compliance [23],

\[
\tilde{D}(x,s) = (1/9) \tilde{B}(x,s) + (1/3) \tilde{J}(x,s),
\]

where

\[
\tilde{e}(x,s) = s \tilde{D}(x,s) \tilde{\sigma}_{11}(x,s)
\]

The literature on creep measurements in collagen [25] and gelatin gels [26,27] provides guidance on modeling compliance. A generalized Voigt model is often useful [23,28]

\[
s \tilde{D}(x,s) = D_0 + \sum_{\ell=1}^L \frac{D_{\ell}}{1 + sT_{\ell}} + \frac{1}{s\eta_0}
\]

Constants \( D_\ell \) are compressive creep compliances, and \( T_\ell \) are discrete retardation times that are proportional to viscosity coefficients \( \eta_0 \) of the \( \ell \)th viscoelastic component: \( T_\ell = D_\ell / \eta_0 \). If we can eliminate the last term in Eq. (6) and let \( T_\ell \) be the largest time constant, the Fourier transform of compliance will exist because the region of convergence, i.e., \( s > -1/T_L \), includes the imaginary axis. Equation (6) implies a time-independent elastic strain and \( L \) distinct viscoelastic strains that delay in time the full response. The last term describes the steady-state compressive-flow viscosity coefficient \( \eta_0 \). In weakly compressed tissues, \( \eta_0 \) may represent flow of vascular fluids; in hydrogels it represents movement of unbound water.

A constant uniaxial force \( F_1 \) is suddenly applied at \( t = t_0 \) to a cubic sample of side-area \( A \) along the \( x_1 \) axis. Then \( \sigma_{11}(x,t) = \sigma_{1}(x)u(t-t_0) \), where \( \sigma_{1}(x) = F_1/A \) for the volume element located at \( x \), and the step function \( u(t-t_0) \) is zero for \( t < t_0 \) and one for \( t \geq t_0 \). The Laplace transform of the step stress stimulus is

\[
\tilde{\sigma}_{11}(x,s) = \sigma_{1}(x)s
\]

Combining Eqs. (5)–(7) and taking the inverse Laplace transform yields for \( t > t_0 \)

\[
\epsilon(x,t) = \epsilon_0(x) + \sum_{\ell=1}^L \epsilon_\ell(x) \left[ 1 - \exp\left[ - (t-t_0)/T_\ell(x) \right] \right] + (t-t_0) \sigma_{1}(x) / \eta_0(x)
\]

where strain amplitudes \( \epsilon_\ell(x) = \sigma_{1\ell}(x) / D_\ell \) for \( 0 \leq \ell \leq L \). The strain response of the Voigt model to a step load in time has three components.

The initial elastic response occurs immediately after compression, i.e., \( \epsilon(x,t_0) = \epsilon_0(x) \), before the viscous mechanisms have time to engage. Purely elastic responses are implicitly assumed in “static” elastography techniques that ignore time-varying strain [24,29,30]. If \( \sigma_{1}(x) = \sigma_{1} \) is constant throughout the volume, then the instantaneous elastic response is directly proportional to the compressive compliance \( D_0 \) (and inversely proportional to the elastic modulus \( E_0 \)) in the volume element. Stresses in heterogeneous media, whose volume elements have unknown boundary conditions, vary unpredictably with position. Strain images in such media must be carefully interpreted to infer stiffness.

The second term defines the time-varying viscoelastic (VE) response: \( \epsilon_{VE}(x,t) = \epsilon(x,t) - \epsilon_0(x) - (t-t_0) \sigma_{1}(x)/\eta_0(x) \). In solids, strain builds exponentially over time with rate constants \( T_\ell \) until the total strain reaches the steady-state value \( \epsilon(x) = \sigma_{1}(x)/\eta_0(x) \) at \( t > T_L(x) \). Measurable viscoelastic responses are from breakage and reformation of weak molecular bonds, release of polymer filament entanglements [28], and other internal restructuring.

The third term in Eq. (8), which varies linearly in time, describes viscous flow within the polymer; e.g., curve a in Fig. 1(a). If time-varying strain plateaus (curve b), the polymer behaves as a solid. VE solids are modeled with Eq. (8) by setting the last term to zero. Model parameters \( D_\ell, T_\ell, \) and \( \eta_0 \) that vary spatially are candidate parameters for diagnostic imaging. Because \( \sigma_{1}(x) \) is unknown in practice, we study \( \epsilon_\ell(x) = D_\ell \sigma_{1}(x) / \eta_0(x) \) in place of \( D_\ell \).

Ultimately, the value of \( \epsilon_\ell, T_\ell, \) and \( \eta_0 \) as diagnostic imaging features depends on their sensitivity and specificity to disease-related changes in tissue structure and biochemistry [6]. The discrete compliance model of Eq. (6) is attractive because it offers a testable number of parameters that may be interpreted in terms of polymer structure. Fung [21] and others warn against determining
the order of the model by blindly fitting model functions to data. The retardation spectrum [28,31] described below provides another tool for estimating retardance time distributions.

First, we examine the Fourier spectrum of the VE creep response in two ways. One describes the spectrum of the creep measurement $\tilde{e}_{VE}(\omega)$ to determine sampling requirements. Strain is sampled in time at the frame rate of the ultrasound system. Another describes the frequency spectrum of the medium response $\tilde{D}(\omega)$.

**Fourier Spectra.** The Fourier transform of the VE response to a uniaxial step stress may be found from the Laplace domain representation of Eqs. (5)–(7) by substituting $s = i\omega$

$$\tilde{e}_{VE}(\omega) = \frac{1}{i\omega} \sum_{i=1}^{L} \frac{1 - i\omega T_i}{1 + i\omega^2 T_i^2}$$

Equation (9) shows the Nyquist frequency to be $f_N = \omega_L/2\pi = 1.5$ Hz, requiring a frame rate of at least 3 Hz to faithfully record creep with $T_i \approx 3$ s. To visualize the lowest frequency peak at $\omega_L$, in this example, corresponding to $T_i = 100$ s, the acquisition time should be $(2\pi)/\omega_L > 628$ s, preferably longer. Acquiring data for shorter times truncates the spectrum at low frequencies without distorting higher frequency values, but creates difficulties in determining model order from data as described below. In vivo breast imaging techniques allow patient acquisition times between 20 and 200 s. Acquisitions in hydrogel samples are often on the order of 2500 s.

The two peaks in the frequency spectrum arise from $L=2$ roots (nonzero poles of Eq. (6)) at $s = -1/T_i$; both are real and negative. They correspond to spectral peaks at $\omega_2 = 1/T_1$ [33], of height $D_2/2$, and $-6$ dB peak width $\Delta\omega_0 = 2\sqrt{3}/T_i$. The latter property shows that $T_i$ must be widely separated to resolve their peaks on the frequency axis. The pole at $s = 0$ from the steady-state viscosity term must be eliminated for the Fourier transform to exist. Poles of the model uniquely determine the time-varying properties of the material.

**Retardation Spectra.** It is attractive to adopt a discrete model for compliance; e.g., Eq. (6). Low-order models with few components that correspond to specific structural and biochemical features yield the diagnostic imaging parameters we seek. However, data from tissues [21] and gels [28] suggest broad continuous distributions of retardance times $\tau$. Schwarzl and Staverman [31] proposed a technique for estimating continuous spectra $L(\tau)$ from creep data. To facilitate direct comparisons with Fourier spectra, we plot $\tilde{D}(\omega) = L(\tau)|_{\omega = \omega_L}$. The two forms of $L$ are rejections of each other about the ordinate, followed by a translation along the logarithmic abscissa.

$L(\tau)$ is introduced by considering the Laplace transform of Eq. (8) for a step stress stimulus and a continuous distribution of compliance

$$\tilde{D}(x,s) = \frac{D_0(x)}{s} + \int_0^\infty d\tau \frac{D_i(x,\tau)}{s(1 + s\tau)} + \frac{1}{s^2 \eta_i(x)}$$

Equation (11) where $D_i(x,\tau)$ is the sampled compliance function obtained when the discrete sum is converted into an integral as shown in [34]. Substituting $L(\tau) = \tau D_i(x,\tau)$ and noting that $d\ln s$ in $d\ln s/d\tau = 1$ and $\tau = \tau(x)$, the inverse Laplace transform of Eq. (11) for $t > t_0$ is

$$D(x,t) = D_0(x) + \int_{t_0}^t d\tau \int_0^\infty d\tau L(x,\tau) \left[1 - \exp\left(-(t - t_0)\tau(x)\right)\right] + \frac{(t-t_0)}{\eta_i(x)}$$

Equation (12), A method for estimating $L$ from creep compliance estimates $\tilde{D}_{VE}$ was described by Tschoegl [23]:
describe the input distribution of \( k \) inversely, retardation spectral estimates approach the input distribution, placing greater emphasis on filter design. However, estimates become unstable as \( D \) saturated noiseless creep data assuming a log-normal input distribution, while the curves are bandwidths for log-normal distributions. The abscissa is \( \tau \) from the log-normal input distribution \( \tilde{D}(t) \) for \( k = 1, 2, 5, 6 \), are compared to the Fourier spectrum (FS) \( \tilde{D}(\omega) \), computed from the same data. (b) \( L^k(\omega) \) estimates without noise in the creep data and with noise (signal-to-noise ratio = 32.2 dB). A ninth-order polynomial filter was applied to the noisy data before estimation.

2.3 Shear Stress and Strain. The unconfined boundaries of arbitrarily shaped, heterogeneous media subjected to uniaxial stress stimuli in imaging experiments can violate the assumptions leading to Eq. (3). To study the effects, we compare parameters from the carefully controlled geometry of standard rheometer measurements to those from creep imaging experiments. Our interest is with average properties, so the positional dependence is ignored for these nonimaging measurements.

The constitutive equation is calculated in the Laplace domain from Eq. (3)

\[
\overline{\tau}_{12}(s) = \frac{1}{2} \tau \tilde{\eta}(s) \overline{\tau}_{12}(s)
\]

Bulk compliance terms are negligible in rotational shear measurements. For a step shear stress, \( \overline{\tau}_{12} = \sigma \overline{J}(t-t_0) \), and assuming the

The effects of measurement noise are shown in Fig. 2(b). Adding white Gaussian noise with signal-to-noise ratio 32.2 dB (typical of rheometer data described below) introduces bias particularly at high frequency. Figure 3 predicts the amount of bias introduced as the width of the log-normal input distribution increases. The data suggest that a 150 s bandwidth can be estimated with acceptable bias by a sixth-order estimate \( L^{(6)}(\omega) \).
Voigt model in shear, the observed creep in the time domain is

\[ \gamma_2(t) = \gamma_{120} + \sum_{m=1}^{M} \gamma_{12,m} \left[ 1 - \exp\left( - (t - t_0)/T_m \right) \right] + (t - t_0) \alpha' / \eta_0 \text{ for } t > t_0 \]  

Measurable shear creep is related to the corresponding strain tensor via \( \gamma_2 = 2 \varepsilon_2 \) [23]. In addition, \( \gamma_{12,m} = \alpha' \eta_m \) for \( 0 \leq m \leq M \) and \( \eta_0 \) is the steady-state shear-flow viscosity coefficient. To account for the geometry of the cone-plate viscometer, the ratio \( \gamma_2(0)/\gamma_{12}(t) = \varphi / \Gamma \), where \( \varphi \) is the angular displacement, \( \Gamma \) is the applied torque, and \( \Lambda = 2 \pi R^3 / 3 \) is a geometric factor that depends on the radius of the cone (\( R \approx 30 \) mm) and on the angle between the cone and plate (\( \phi \approx 4 \) deg).

Compression and shear measurements may be compared through Eqs. (8) and (16). Compressive and shear creep compliances are, respectively

\[ D(t) = \varepsilon_1 / \sigma_0 = D_0 + \sum_{m} D_m \left[ 1 - \exp\left( - t'/T_m \right) \right] + t' / \eta_0 \]

\[ J(t) = \gamma_2 / \sigma_0 = J_0 + \sum_{m} J_m \left[ 1 - \exp\left( - t'/T_m \right) \right] + t' / \eta_0 \]  

for \( t' = t - t_0 \geq 0 \). From Eqs. (4) and (5) we have \( D(t) = J(t)/\Gamma + B(t)/\eta_0 \). Thus, model parameters for the two experiments may be compared directly only for “incompressible media” where bulk compliance \( B(t) \) is negligible. Bulk compliance can be related to compressive compliance and Poisson’s ratio \( \nu \) in the Laplace domain:

\[ s \tilde{B}(s) = 3s \tilde{D}(s) \left( 1 - 2s \tilde{\kappa}(s) \right), \quad \tilde{\kappa}(s) = -\tilde{\kappa}_2(s) / \tilde{\varepsilon}_1(s) \]

We can then use limit theorems [23] to find in the time domain \( \tilde{B}(s) \approx 3 \tilde{D}(s) (1 - 2 \kappa(s)) \) at \( s \to 0 \) and \( \tilde{B}(0) = 3 \tilde{D}(0) (1 - 2 \kappa(0)) \) at \( s \to \infty \).

### 2.4 Uniaxial Compressive Strain: Stress Relaxation and Relaxation Spectra

Stress relaxation experiments are conducted in which samples are stimulated with a uniaxial step strain while stress is measured over time. This nonimaging technique provides spectral data under confined boundary conditions that could not be obtained using creep measurements with our instruments.

Analogous to Eq. (3), the generalized viscoelastic Hooke’s law for stress relaxation is [23]

\[ \tilde{\sigma}_j(s) = \begin{pmatrix} \tilde{s} \tilde{K}(s) - \frac{2}{3} \tilde{G}(s) \end{pmatrix} \tilde{\delta}_j + 2s \tilde{G}(s) \tilde{\varepsilon}_j(s) \]

where \( \tilde{\delta}_j = \tilde{\varepsilon}_1(s) + \tilde{\varepsilon}_2(s) + \tilde{\varepsilon}_3(s) \), \( \tilde{G}(s) \) and \( \tilde{K}(s) \) are shear and bulk moduli, respectively; they are analogous to the compliances \( \tilde{J}(s) \) and \( \tilde{B}(s) \) measured in creep. If the sample boundaries are confined in the manner described in Method B below, then there is only one nonzero strain tensor

\[ \tilde{\sigma}_{ii}(s) = \begin{pmatrix} \tilde{s} \tilde{K}(s) + \frac{4}{3} \tilde{G}(s) \end{pmatrix} \tilde{\varepsilon}_{ii}(s) = s \tilde{M}(s) \tilde{\varepsilon}_{ii}(s) \]

Equation (19) relates the measurable compressive longitudinal wave modulus \( \tilde{M} \) for the confined sample to fundamental relaxation moduli \( \tilde{K} \) and \( \tilde{G} \) in the Laplace domain [23].

Alfrey’s rules [23] describe how to select a Maxwell model for \( \tilde{M}(s) \) that is conjugate to the Voigt model of Eq. (6): \( s \tilde{M}(s) = M_0 + \sum \alpha M_1 s^{1/2} (1 + s t_0) \). Applying a uniaxial step strain stimulus, \( \varepsilon_{11}(t) = \varepsilon_{11}(t - t_0) \), the time-varying wave modulus is

\[ M(t) = M_0 + \sum \alpha M_1 \exp(- t'/T_n) \text{ for } t' = t - t_0 > 0 \]  

where \( T_n \) are discrete relaxation time constants. Unfortunately, it is not easy to relate \( T_n \) directly to retardation time constants \( T_i \) for this geometry.

If the sample boundaries are unconfined, all three strain tensors are nonzero. The axial stress tensor is

\[ \tilde{\sigma}_{ii}(s) = \frac{9s \tilde{K}(s)}{3s \tilde{K}(s) + s \tilde{G}(s)} \tilde{e}(s) \tilde{\varepsilon}_{ii}(s) \]

Applying the same step strain stimulus, the compressive relaxation modulus is

\[ E(t) = \frac{\varepsilon_{11}(t)}{\sigma_0} = \sum_{i=1}^{N} E_i \exp(- t'/T_i) \text{ for } t' = t - t_0 > 0 \]

\( E(t) \) may be compared to creep compliance \( D(t) \) in the Laplace domain by \( E(s) D(s) = \sigma^2 \). Alternatively, \( \int_0^t E(t) D(t) d \tau = t \), suggesting \( D(t) E(t) \approx 1 \) [28]. When \( \nu = 0.3 \), \( D(t) E(t) = 1 = \tilde{G}(t) / \tilde{D}(t) \). From Eq. (22), the elastic (Young’s) modulus is defined as \( E_0 \equiv \sigma_{11}(t_0)/\varepsilon_0 = \sum_{i} E_i \).

Similar to the methods described in Sec. 2.2 for retardation spectra, relaxation spectra \( H(\tau) \) and \( H(\omega) \) may be estimated from stress relaxation data [23,28]. \( H(\tau) \) is the distribution of relaxation times that determines the time dependence of a modulus. For confined samples, a continuous distribution of relaxation times is modeled as \( \tilde{M}(s) = M_0 \int_0^\infty d \tau \exp(-s \tau) H(\tau) \) [23], and similarly for \( H(\tau) \). Depending on context, \( H(\tau) \) refers to either \( H_M \) or \( H_E \).

### 2.5 Gelatin Model

The above set of measurement parameters was explored by selecting animal-hide gelatin hydrogels for experimentation. Gelatin gels have an extensive literature of mechanical measurements [26–28,35–39], are simple to construct, are elastically uniform within the resolution of the ultrasonic imaging system, and manifest essential tissue-like material features.

At room temperature and pressure, gelatin gels are lightly cross-linked amorphous polymers surrounded by layers of structured water. Depending on the stress stimulus, the strain response can have both solid and fluidic features. The peptide structure and molecular surface charges determine the viscoelastic behavior; consequently, the properties vary with pH, salt concentration, and thermal and mechanical histories. Gelatin gels have lower material strength than the connective tissues from which they derive because the collagen is denatured. Chemical and thermal stresses that break down the natural Type I collagen super-structure during processing is only partially reconstituted during gelation and with many fewer covalent bonds [40]. While fragments of the original triple \( \alpha \)-helix structure reform, most of the protein molecules remain as peptide chains that are randomly tangled among the sparse helical fragments (see Fig. 4 from [38]). The molecular weight of the protein molecules is generally above 125 kDa, suggesting a matrix of relatively long and interconnected peptide chains. Unlike natural connective tissue collagen, there is no polysaccharide gel surrounding these chains [41]. Yet there are many reactive ionic groups exposed that adsorb water molecules.

Desiccated gels retain about 10% water that is tightly bound to the charged residues. In this role, water forms stabilizing intramolecular hydrogen bonds [38]. Increasing hydration adds layers of water molecules more viscous than free water because of its polar attraction to the charged protein backbone [42]. Near the highest hydration levels that still yield, structured water layers are added with increasingly weaker binding forces. The outermost layers remain bound under a load if the resistance to flow \( \eta_0 \) is greater than the applied forces \( \sigma_0 \). From Eqs. (8) and (16), gels...
may be considered VE solids when $\sigma_d/\eta_r \ll 1$ (curve b in Fig. 1(a)). Otherwise, they exhibit the viscous flow of rheodic materials (curve a).

Gelation is initiated within molten gelatin near sites of the randomly located $\alpha$-helices [38]. When the temperature falls below about 30°C, polymerization is nucleated, and aggregates of hydrogen-bonded protein molecules form. Material strength increases with gelatin concentration because the aggregate bond density increases. Hydrogen bonds, which break and re-form under a load, are a source of viscoelastic creep, i.e., $\varepsilon_V(t)$. The distribution of adhesive force strengths in the polymer determines the retardation spectrum. Covalent bonding among sparse helical fibrils [43] as well as the strong intra-molecular bonds both contribute to the initial elastic response $\varepsilon_0$. The covalent-bond density can be increased to stiffen gels by adding aldehydes [37]. Thus, melting temperature is increased and temporal stability improved, as is required for tissue-like imaging phantoms.

2.6 Gelatin Sample Preparation. To each 100 ml of deionized water, we add 13 ml of n-propanol and 6.5 g (12.4 g) of 275-bloom, animal-hide gelatin (Fisher Scientific, Chicago, IL) to arrive at a 5.5% (10%) gelatin concentration. The solution is heated at 60°C until visually clear (~30 min) before adding 0.3 ml of formaldehyde (37% w/w). The hot solution is poured into a rigid container and quiescently cooled. Although gelatin congeals in hours, it continues to cross-link for many days. Samples are stored at room temperature 1–5 days before conducting measurements. The elastic modulus $E_0$ of gelatin is known to increase linearly with log(time) [44]. "Stiff" samples are 10% gelatin by weight and "soft" samples are 5.5% gelatin; both are above the critical gelation concentration [45]. Since $E_0$ is proportional to the square of gelatin concentration [44,45], 10% gels are roughly three times stiffer than the 5.5% gels.

Samples made for compression measurements are either 5-cm cubes or cylinders of diameter 15 mm and height 15 mm (via 10-cc syringes). Cubic gel samples are removed from the molds before measurement to free the boundaries from confinement. Cylindrical gel samples remain in the syringe as uniaxial compressions are applied under confined boundary conditions using the syringe piston. Shear measurements are made on samples formed in the rheometer as described in the next section. Indenter measurements are made near the axis of cylindrical samples of diameter 60 mm and height 6 mm that are removed from their containers.

Two types of commercially available gelatin are studied. Type A gelatin (pH 6) involves acid processing of collagen-rich media, whereas Type B gelatin (pH 5) is obtained from alkaline processing. Type A preserves more of the natural collagen structure but contains impurities that affect mechanical properties. Type B gelatin is a purer form of collagen molecule, yet it undergoes greater denaturation so that fewer fibrils re-form, and the reconstituted structure is less similar to native connective tissues.

2.7 Viscoelastic Measurement Techniques. All gelatin measurements are made at ambient room temperature and pressure.

Method A: Uniaxial Compression in Unconfined Samples. A flat plate compresses the top surface of a cubic gel sample downward as the sample rests on a digital force balance (Denver Instruments Co., Model TR-6101, Denver CO) (see Fig. 5(a)). A motion controller (Galil Inc., Rocklin CA) is programmed to apply a small preload to establish contact. A short-duration (~1 s) ramp stress is then applied along the direction normal to the sample surface to initiate creep measurements. The final force is held constant over time by using the balance output as feedback. Sampling the balance output at 3.4 samples/s, the motion controller adjusts the compressor position within 0.1 $\mu$m so the applied force remains constant during the experiment as the sample creeps. The position of the compressor indicates displacement for creep estimates. The effects of the ramp stimulus relative to a step stimulus are discussed in Appendix A.

For a cubic sample of height $h$, we measure displacement $\Delta h$ and force $f[N]=\text{mass}[\text{kg}] \times 9.81$. These quantities are converted to true stress $\Gamma = \Delta h/h F/A_0 [\text{Pa}]$ and true strain $\ln(1 + \Delta h/h)$, where $A_0$ is the unloaded sample area contacting the balance. Mineral oil is applied to all exposed sample surfaces to minimize desiccation and to allow boundaries to freely slip under a load.

Several gelatin samples are constructed from each preparation. If a sample is used more than once to repeat an experiment, we follow the rule of resting samples more than twice the acquisition time of the previous experiment (Appendix B). A typical creep acquisition is 2500 s. Those samples are rested 2 h between measurements.

Method A is often used to acquire time sequences of axial strain images $\varepsilon_{ij}(x,t)$ by flush mounting a linear array transducer into the compression plate [46], as shown in Fig. 5(a). We can also apply strain stimuli to estimate stress relaxation and complex compliance/modulus parameters, or we can modify the technique to estimate lateral strain for Poisson’s ratio estimates. In the latter case, samples are submerged in a water-alcohol solution without the force balance, and a step strain $\varepsilon_{1l}(t) = \varepsilon_u(t-t_0)$ is applied. The transducer in Fig. 5(a) is rotated 90 deg to scan the sample from the side and measure true lateral strain $\varepsilon_2(t) = \ln(1 + \Delta w(t)/w)$. A sample of width $w$ will expand a time-varying distance $\Delta w(t)$ when compressed from above and held. Therefore, Poisson’s ratio is $\nu(t) = -\ln[1 + \Delta w(t)/w]/\ln(1 + \varepsilon_u)$ for $t > t_0$.

Method B: Uniaxial Compression in Confined Samples. Method B is illustrated in Fig. 5(b). Cylindricaly shaped samples encased in rigid plastic are compressed uniaxially with a step strain to measure stress relaxation. There is a porous bottom surface that allows fluids to pass but not the gelatin. After preparation in a sealed syringe, the end is removed and a moist gauze and fine screen are attached to the expose gelatin surface before mounting.

Fig. 4 Illustration of collagen structures in connective tissue (fibril) and in gelatin (aggregates)
A 1 s compressive ramp displacement is applied from above with the motion controller and held constant while measuring the force. Displacement and force are converted to true stress and strain as shown above. Equation (20) describes the wave modulus for confined samples.

Originally the goal was to measure creep in confined samples. However, Method B apparatus is unable to generate artifact-free creep data, so we settled for stress relaxation data. Comparisons are made using the analysis in Sec. 2.4 and are discussed below.

**Method C: Cone-Plate Rheometer Measurements.** Method C is illustrated in Fig. 5(c). We measured shear compliance under the strict boundary conditions of a Haake cone-plate rheometer (Thermo Electron Corp., Model RS150, Waltham MA) to validate compressive compliance estimates. Comparisons were made by applying the analysis of Sec. 2.3.

Molten gelatin was poured into the cone-plate rheometer plate at approximately 30°C so that it covered the edges of the cone. The sample was closed to outside air and cured 1–4 days before measurements. This preparation eliminated slippage at surfaces when the sample was sheared. A short duration ramp shear stress at either \(\sigma^1 = 3\) or 30 Pa was applied and held while strain was recorded for times up to 3000 s at a rate of 3 samples/s. Equation (16) represents data acquired by these measurements. The cone-plate rheometer was also capable of harmonic stimuli at frequencies between 0.0001 and 15 Hz.

**Method D: Indentation Methods.** Method D is illustrated in Fig. 5(d). Indentation is a widely accepted method for estimating the elastic modulus. Gel samples, each 60 mm in diameter, were prepared and stained with a known sinusoidal displacement stimulus at a frequency of 0.02 mm/s while measuring the applied force on the balance. Ten cycles were recorded for each sample at three surface frequencies.

2.8 Data Processing. The digital balance samples force with a variable time interval due to limitations of the instrument. However, a time stamp for each sample is available. The average sampling frequency is 3-4 samples/s. Creep data are interpolated to 10 samples/s and then downsampled a factor of 5 to facilitate curve fitting; the final sampling interval is \(\Delta t = 0.5\) s.

VE parameters are estimated by fitting creep data, e.g., curve b in Fig. 1(a), to an \(L\)-th-order Voigt model, where \(L=1, 2\) or 3. Fitting is achieved using optimization techniques using MATLAB’s Optimization Toolbox LSQCURVEFIT, where \(D(t)\) from Eq. (17) is the function that is fit to the measurements \(\tilde{D}[n\Delta t] = \tilde{C}[n\Delta t]/\sigma_0\). The unbounded Levenberg-Marquardt optimization option is selected. Monte Carlo tests showed that the algorithm quickly converges if the initialization parameters are close to the true values and the number of fit parameters is minimized.

We first estimate steady-flow viscosity in a preprocessing step so it can be subtracted from the data before curve fitting. The estimate \(\hat{\eta}_0\) is found by computing the derivative \(\hat{\dot{D}}[t] = (d\ddot{C}/dt)/\sigma_0\) over the measurement time, identifying the time at which \(\hat{\dot{D}}(t)\) becomes constant with time, and then averaging subsequent values: \(\hat{\eta}_0 = \frac{\sum \hat{\dot{D}}[n\Delta t]/N_\Delta}{N_\Delta}\) for the \(N_\Delta\) points, where \(t = n\Delta t > 2T_{\text{max}}\). Eliminating the steady-state viscosity term before model fitting speeds convergence.

2.9 Goodness of Fit and Model Order. Results from fitting \(N\) preprocessed creep compliance data points to a Voigt model of order \(L\) with fit parameters \(\theta = (\varepsilon_0, \varepsilon_1, T_1, \ldots, \varepsilon_i, T_L)\) are evaluated by computing the \(\chi^2\) value [48]

\[
\chi^2 = \sum_{n=1}^{N'} \left( \frac{(\hat{D}[n\Delta t] - D[n\Delta t; \theta])^2}{\text{var}_D} \right)
\]

For a third-order model, \(\theta\) has dimension \(2L+1 = 7\). In addition, \(\text{var}_D\) is the variance of \(\hat{D}(t)\) estimates. \(\chi^2\) has \(\xi = N' - (2L+1)\) degrees of freedom. We compute the probability \(Q(x^2; \xi)\) that the observed chi-square exceeds \(x^2\) by chance assuming the measurement errors are normally distributed. \(Q(x^2; \xi)\) were computed using the incomplete gamma function [48]. We select \(L\) by finding the lowest-order model for which \(Q > 0.2\). Curve fitting in the time domain favors long response times, so \(Q\) plays an essential role in helping us determine model order.

3 Results

3.1 Viscosity. Shear creep experiments (Method C) were conducted to estimate the steady-state shearflow viscosity coefficient \(\eta_0\). Figure 6(a) shows there is a constant equilibrium strain for the
step stress amplitude $\sigma'_1 = 3$ Pa, indicating no fluid flow. However, there is a linearly increasing strain in the same samples for the $\sigma'_1 = 30$ Pa stimulus, indicating that flow occurs. Using the 30 Pa data, we estimate viscosity versus time in Fig. 6 to find the steady-state value of $\eta'_0 \approx 10^7$ Pas for Type B gelatin. A creep recovery method was also applied (Fig. 6(c)), to Type A gelatin (5.5%) at 100 Pa shear stress to find $\eta'_0 \approx 10^8$. Estimates from the creep and recovery phases of Fig. 6(c) are approximately equal as expected.

Gelatin gels are rheodictic only when sufficiently stressed. They behave like a VE solid ($\eta'_0 \to \infty$) at 3 Pa and like a VE polymer saturated in a viscous fluid at stresses above 30 Pa. Viscosity measurements in gelatin gels are constant above a stress threshold, although the values depend on gelatin concentration and type. A power-law dependence of $\eta'_0$ on gelatin concentration has been observed by others [45].

### 3.2 Linearity

Unconfined gelatin samples were strained uniaxially with the harmonic stimulus $\varepsilon(t) = \varepsilon_0 \sin(\omega_0 t)$, where $\omega_0 = 2\pi \times 0.03 \text{ mm/s}$, to generate the stress-strain curves of Fig. 7(a). Data shown are from the ninth cycle. Considering strain above 0.01, the on-load halves of each curve (top lines) are linear.
with a correlation coefficient $r^2=0.9999$ for stresses up to 0.86 KPa for the soft gel and up to 3 KPa for the stiff gel. As expected for linear media, no significant change in response times or retardance or relaxation was observed at these stress levels.

To examine linearity in shear, we measured shear creep spectra at $\sigma_0^s=3$ and 30 Pa using Method C. The 3 Pa spectral values were multiplied by 10 and plotted with the 30 Pa spectrum in Fig. 7(c). Visual agreement between the two curves indicates a linear VE creep response in this shear stress range despite the higher noise levels in the 3 Pa data.

3.3 Poisson’s Ratio. Applying the step strain $\varepsilon_{11}(t)=\varepsilon_u u(t-t_0)$ to a 5.5% gelatin cube and measuring $\varepsilon_{22}(t)$ across the entire sample width, we estimated $\nu(t)$ as described for Method A in Sec. 2.7. The results are shown in Fig. 8. Initially, the sample responds incompressibly; i.e., the $\nu(0)=0.5$ within the measurements uncertainty. Within 100 s, however, $\nu(t)$ has fallen to an equilibrium value of 0.473, such that the ratio of equilibrium bulk and compressive compliances increases from zero to $B/\theta/\theta^c = 3[1 - 2\nu(\infty)] = 0.162$. Consequently, creep model parameters obtained in compression and those in shear cannot be directly compared.

3.4 Effects of Acquisition Time. The longest duration response time determines the total required acquisition time. In gelatin gels, data must be acquired up to an hour to visualize the entire bandwidth. However, as acquisitions lengthen, the importance of eliminating the steady-state viscosity term increases. We summarize in Fig. 9 the effects of acquisition time on contrast and retardance time estimates with and without eliminating the viscosity term. Results suggest that the acquisition time must exceed twice the value of the longest response time constant. Failure to eliminate even the weak viscosity term of these gelatin gels introduces bias. Furthermore, decreased acquisition times causes a decrease in contrast.

![Fig. 8 Poisson’s ratio estimates versus time, i.e., $\nu(t)$. Error bars denote one standard deviation computed by propagating displacement measurement errors.](image)

![Fig. 9 (a) Dependence of $T_1$ on acquisition time, and the effect of eliminating steady-state viscosity (linear term in Eq. (17)). $T_1$ and $T_2$ estimates for a third-order Voigt model are shown. (b) Variation of $T_1$ contrast over acquisition time is shown.](image)

![Fig. 10 Comparisons of measurements made using different methods. Samples were all type A gelatin aged three days. (a) Elastic modulus, (b) equilibrium compliance, and (c) steady-state viscosity under compression. Error bars are standard deviations that indicate uncertainty between repeated measurements.](image)
3.5 Validation. In Fig. 10, measurements from different experimental geometries are compared. One of the advantages of using standard rheological models is the opportunity to interconvert some parameters from one experiment into another. Elastic modulus estimates $E_0$ measured using five techniques in compression and shear are plotted in Fig. 10(a): Method A with step stress (CR), step strain (SR), and harmonic stress (Osc) stimuli, and Methods C and D. Mean values of $E_0$ agree within 6%. Figures 10(b) and 10(c) display estimates of equilibrium compliance and steady-state viscosity from step stress (CR) and strain stimuli of Method A after the response from step strain is converted to an equivalent step stress response (SR $\to$ CR) under the assumption $D(t)E(t) = 1$. No significant differences were found (Student $t$-test; $\alpha = 0.05$).

3.6 Image Contrast. Viscoelastic measurements of gelatin, modeled as third-order discrete processes, are characterized by eight parameters. Which of these parameters are best for imaging? In practice, the answer depends on the conditions and reasons for obtaining the image. Yet, we can illustrate the point by estimating parametric contrast for different gelatin concentrations that simulate conditions of a fibrotic lesion.

For two homogenous phantoms with gelatin concentrations of 5.5% and 10%, the contrast magnitude for parameter $X_1$, $X_2$, and $X_3$ is proportional to the compliance distribution. However, it is well known that stresses in heterogeneous media vary with position [49]. For example, Fig. 11(b) is an $\epsilon_0$ image of a 5.5% gelatin block into which a stiff cylindrical inclusion of 10% gelatin [24] is placed. Strain in the regions surrounding the inclusion vary because the local stresses are nonuniform. $T_1$, and either $T_2$ or $T_3$, depending on available acquisition times, are also reasonable choices to represent fluid and matrix responses of gelatin. An example $T_3$ image is shown in Fig. 11(c). Lesion areas are brighter, indicating that mechanisms take longer due to the increased collagen density when compared to softer background areas. $\epsilon_0$, $T_1$, and $T_2$ are the three parameters currently used for viscoelastic imaging [46]. The measurements of Fig. 11 should be repeated to select parameters for imaging biological tissues.

3.7 Viscoelastic Spectra. Figure 12 displays Fourier spectra with corresponding response time distributions for four experiments. Specifically, we plot $\tilde{\sigma}(\omega)$ and $\tilde{\sigma}^{(3)}(\omega)$ in part (a), $\tilde{M}(\omega)$ and $\tilde{H}^{(3)}(\omega)$ in part (b), $\tilde{J}(\omega)$ and $\tilde{L}^{(3)}(\omega)$ in part (c), and $\tilde{\varepsilon}(\omega)$ and $\tilde{H}^{(3)}(\omega)$ in part (d). The notation $\tilde{L}^{(3)}$ indicates the approximation to Eq. (13) converges at $k = 3$. The Fourier spectral bandwidth in each case is less than 10 rad/s.

Table 1 lists parameters estimated by curve fitting the data in Fig. 12 to model functions. The $\chi^2$ probabilities in Table 1 show that a third-order model is required for uniaxial compression (Figs. 12(a), 12(b)) to meet the goodness-of-fit criteria for accept-

Table 1: Viscoelastic parameters for 5.5% gelatin acquired by fitting measurements to model functions. First column lists the discrete viscoelastic model order. Second column contains compressive compliance [kPa$^{-1}$] and relaxation time [s] constants from data of Fig. 12(a). Third column contains wave modulus [kPa] and relaxation time [s] constants from Fig. 12(b). Fourth column contains shear compliance [kPa$^{-1}$] and retardance time constants from Fig. 12(c). Fifth column contains compressive relaxation modulus [kPa] and relaxation time constants from Fig. 12(d). $Q$ is the probability of the goodness-of-fit test.

<table>
<thead>
<tr>
<th>MO</th>
<th>Fig. 12(a)</th>
<th>Fig. 12(b)</th>
<th>Fig. 12(c)</th>
<th>Fig. 12(d)</th>
</tr>
</thead>
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<tr>
<td>2</td>
<td>$D_0=0.109$</td>
<td>$M_0=307$</td>
<td>$J_0=0.908$</td>
<td>$E_0=0.46$</td>
</tr>
<tr>
<td></td>
<td>$D_0=0.005$</td>
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<td>$T_1=13.7$</td>
<td>$T_1=22$</td>
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<td>$D_1=0.006$</td>
<td>$T_2=338$</td>
<td>$T_2=198$</td>
<td>$T_2=302$</td>
</tr>
<tr>
<td></td>
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<td>$Q=0$</td>
<td>$Q=0.34$</td>
<td>$Q=0$</td>
</tr>
<tr>
<td>3</td>
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<td>$M_0=307$</td>
<td>$J_0=0.904$</td>
<td>$E_1=0.43$</td>
</tr>
<tr>
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<td>$D_0=0.004$</td>
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<tr>
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<td>$Q=0.48$</td>
<td>$Q=0.41$</td>
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</table>

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constants.

The spectrum of the sheared sample (Fig. 12) is very similar to the stress relaxation response of Fig. 12. Ferry refers to this as "entanglement coupling." Because these movements occur over a large spatial scale, longer response delays are expected.

Tschoegel [23] also addresses the dynamic behavior of weakly cross-linked polymers such as gelatin gels. He refers to it as pseudo-arheoditic because the frictional forces between matrix fibers that retard strain in creep experiments can appear as delayed fluid flow. When the magnitude of frictional forces varies over time, a portion of the VE response is delayed, which generates a bimodal spectrum. The Ferry and Tschoegel descriptions are consistent if one considers that the time required for fibers to be straightened before they are dragged through the matrix could be the source of the characteristic delay. In that case, spectral peak frequencies are expected to depend on the molecular weight and surface charge density of the matrix fibers. A working hypothesis for biological tissues is that disease states alter properties of the extracellular matrix—the natural polymer of the body—to generate disease-specific contrast in images of viscoelastic parameters.

In both short-duration (fluid) and long-duration (matrix) responses of gelatin, frictional forces from bending peptide chains and their attraction to the surrounding structured fluids vary in strength given the randomness of the matrix geometry.
Thus, there are not two response times as expected from discrete modeling but two distributions of times as observed from the spectra of Figs. 12(a), 12(b), and 12(d). The observation that \(L^k\) and \(H^k\) were found to converge suggests continuous distributions of response times are reasonable to assume.

Confining samples as in Fig. 12(b) forces fluids to flow before the matrix can respond [50]. In the unconfined samples of Figs. 12(a) and 12(d); however, these processes can begin simultaneously. We see from Table 1 that response times for the high-frequency peak, i.e., 3.5 and 5.5 s for the unconfined samples, decreases to 1.5 s in the confined sample, while changes in the low-frequency response times are less pronounced. Sample confinement appears to separate and narrow the distributions, as expected from the Ferry and Tszoege descriptions.

In shear creep (Fig. 12(c)), tensile forces are applied to the matrix instead of compression. Forces on the matrix fibers near the circumference of the cone-plate are much larger than those near the center of rotation. Consequently, even small rotations engage the matrix immediately. Since polymers resist tensile deformations more than comparable compressive deformations, the larger low-frequency matrix response observed compared to the high-frequency fluid response is expected. Thus, the skewed, unimodal appearance of the spectrum in Fig. 12(c) may reflect an increased relative weighting of the low-frequency response.

Clearly, low-order discrete viscoelastic models do not provide physical descriptions of polymers. Rather they are parsimonious summaries that help guide selection of imaging parameters. Our burden is to show those parameters are related to essential biological processes. We are concerned that apparently bimodal spectra require third-order discrete models to meet the \(\chi^2\) criteria. At this time, we recommend using spectra to observe the number of modes, and then averaging time constants detected within each mode. For the spectra of Fig. 12(a), where the \(\chi^2\) criterion suggests a third-order model, we would nevertheless average time constants corresponding to the two lowest-frequency poles and therefore report \(T_1=5.5\) s and \(T_2=209.5\) s.

Given the interpretation above, it seems that images of elastic strain \(\varepsilon_0\) and the retardance times \(T_1\) and \(T_2\) form a concise feature space for strain imaging investigations. The frame rate of current ultrasound systems easily provides sufficient temporal resolution to sample the viscoelastic response bandwidth without aliasing. The challenge for viscoelastic parameters is to acquire data over a sufficient time duration to sample the low-frequency spectral response and estimate steady-state viscosity \(\eta_0\). The longest response time for gelatin is less than 400 s, so acquisitions of 800 s are sufficient when \(\eta_0\) is large. Even though the steady-state viscosity of gelatin is relatively high, it competes with viscoelastic responses and therefore must be eliminated before analyzing the VE response to minimize parameter biases. The threshold for rheodictic strain responses in gelatin is low: less than 30 Pa.

A different approach to viscoelastic modeling that is gaining momentum models the constitutive equation as a fractional derivative [23,51]. Instead of exponential time dependencies, strain retardation (or stress relaxation) is modeled as algebraic decays [52,53]. Mathematically, \(\varepsilon_0(x,t)=D_0(x)\alpha(x,t)^\alpha\), where \(D^\alpha\) is the fractional derivative operator applied to the stimulus and \(0<\alpha<1\). The fractional derivative result can form a concise feature representation by the two parameters: \(D_0(x)\) and \(\alpha(x)\). In fact, [51] shows that there is also a molecular basis for interpreting these parameters in polymer solutions. However, the interpretation in cross-linked polymeric solids with arrhenodictic behavior, such as gelatin and soft connective tissues, is still empirical in the sense that \(\alpha\) is a characteristic parameter not directly connected to molecular structures.

The creep response in gelatin is well represented by linear viscoelastic theory for applied stresses up to 3 kPa, although the range depends on gel stiffness. The literature for some biological tissues shows a lower threshold for nonlinear responses [21]. The question of interpreting VE parameters for images obtained during large, nonlinear deformations is open [54]. Anecdotal evidence from imaging [17,24] shows there is little change in contrast even for large compressions where nonlinear responses are clearly expected. While interpretation of parameters in terms of polymer structure may require linearity, detection of features in imaging based on contrast may not. In addition, strain errors generated by violations of the linear assumption are relatively small compared with other sources of imaging errors. For example, strain variance increases as ultrasonic echo signals decorrelate during complex motions of heterogeneous media and from echo fields undersampled with respect to the bandpass of strain gradients [24]. In addition, strain is not directly proportional to compliance when the boundary conditions generate spatially variable local stresses. The generally large object contrast for many biological imaging tasks [4] and the use of Lagrangian coordinates to estimate strain [55] give images of viscoelastic parameters diagnostic value despite violations of assumptions that permit interpretation of results at the molecular scale.

**Acknowledgment**

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**Appendix A: Ramp and Hold Stress Stimulus**

Consider the first-order Voigt model in shear, i.e., \(\tilde{\varepsilon}(t)=\varepsilon_0+J_1(t+T_1)\), where we assume \(t/T_1\geq1\) over the measurement time [23]. Let us apply a ramp shear stress \(\sigma_2(t)=\sigma_2(t_0,t_1)\) over the time interval \((t_0,t_1)\)

\[
\sigma_2(t_0,t_1) = \begin{cases} 
0, & t < t_0 \\
\frac{t}{T_1} - t_0, & t_0 < t < t_1 \\
1, & t \geq t_1
\end{cases}
\]

In the Laplace domain, \(\tilde{\sigma}_2(s)=\sigma_2(t_0,t_1)\), where \(t'=t-t_0\). Combining this information with Eq. (15) and taking the inverse Laplace transform yields shear creep for a ramp stress.

\[
\gamma(t) = \gamma(t) = J_0 \sigma_2 + J_1 \sigma_2 \left[ 1 + \frac{T_1}{\gamma(T_1)} \exp\left(-t/T_1\right)\left[ 1 - \exp(t'/T_1) \right] \right]
\]

for \(t \geq t_1\)

In the limit of \(t' \to 0\), we obtain the response to a step stress \(\gamma(t) = \gamma(t_0) \left[ 1 - \exp(-t/T_1) \right] \). This can be extended to higher-order models for a linear system via

\[
\gamma(t) = \gamma(t_0) + \sum_{m=1}^{M} \frac{\gamma(T_m)}{t} \left[ 1 + T_m \exp\left(-t/T_m\right) \right] - \exp(t'/T_m) \]

for \(t \geq t_1\)

Ramp stimuli reduce the magnitude of viscoelastic responses compared to a step stimulus, particularly at high frequencies, but do not bias retardance time estimates. Results for a compressive ramp stress stimulus yield equivalent effects. For example, Fig. 1(b) compares bimodal spectra simulated with step and 1 s ramp stress stimuli.

**Appendix B: Sample Rest Period Analysis**

Whenever possible, parameter uncertainty was estimated using data from identical samples measured once each. Measurements were repeated on the same sample only when necessary. We avoided repeated measurements on the same sample because viscoelastic responses are known to depend on deformation history. The following study tests how the rest time allowed between measurements affected estimates.
Figure 13 summarizes the results of a creep experiment conducted on two gelatin samples with identical properties using Method A, where \( \sigma_0 = 733 \) Pa. Sample I was rested 1 h between the first two measurements and then 2 h between measurements 2 and 3. Sample II was rested 2 h and then 1. Resting 1 h biased retardance times high by as much as a factor of 2. Waiting 2 h reduced biases significantly, although it is clear that the exact deformation sequence is important. It might seem reasonable to recommend 3 or 4-h rests, except that cross-linking also increases deformation sequence is important. It might seem reasonable to recommend 3 or 4-h rests, except that cross-linking also increases retardance times high by as much as a factor of 2. Waiting 2 h and 3. Sample II was rested 2 h and then 1. Resting 1 h biased the first two measurements and then 2 h between measurements 2 and 3. Sample II was rested 2 h and then 1. Resting 1 h biased the first two measurements and then 2 h between measurements 2 and 3.

### References


The loss compliance $D_\ell(o)$ (Eq. (6)) for a first-order Voigt model is $D_\ell(o) = (D_o o T_0)/(1+o^2 T_0^2)$. The spectrum peaks at $o=1/T_1$ as found from $dD_\ell(o)/do=0$.

To go from the discrete model of Eq. (6) to a continuous distribution of retardance times, we assume $\sum_{t=1}^{\infty} D_t/(1+o T_0)$ can be written as a uniformly sampled function $\sum_{t=1}^{n} D_t/(1+n\Delta T)$, where $T_0=n\Delta T$ for some integer value $n$. Using sampling theory, $\sum_{t=1}^{n} D_t/(1+n\Delta T) = \lim_{\Delta T \to 0} \sum_{t=1}^{\infty} D_t/(1+o T_0)$ has units $[\text{Pa}\cdot\text{s}]^{-1}$.


